

1589-Pos Board B433**Probing the role of Cys-78 in dihydropteridine reductase (DHPR) using Raman Spectroscopy**

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Dihydropteridine reductase (DHPR) catalyzes reduction of the unstable quinonoid dihydropteridine to the active tetrahydropteridine form through the oxidation of NADH. DHPR deficiency or any blockage in biosynthesis of tetrahydropteridine results in development of phenylketonuria (PKU), a progressive neurological illness which does not respond to dietary treatment. It has been speculated that DHPR or metabolites associated with it may have antioxidative properties. In another study, DHPR has been shown to have NADH-ferric reductase activity. This activity is postulated to have an important role in dietary iron uptake. There is an emerging role, though mechanistically unclear at this point, for BH4 in maintaining nitric oxide synthase (NOS) activity. NOS produces the signaling agent nitric oxide (•NO) from L-arginine. In the light of these emerging roles there is a growing need to completely understand how DHPR works.

The mechanism of catalysis in DHPR is not fully understood. Previous studies have suggested the involvement of the thiol group of a cysteine residue in DHPR. Using Raman spectroscopic techniques, the effect of the inhibitors/substrate and/or cofactor on the protonation state of the thiol group is investigated. Raman spectroscopy is particularly effective because of a unique peak at ca. 2500 cm⁻¹ due to the S-H stretch and another one at ca. 900 cm⁻¹ due to the C-S bond. Peak shifts and modulation in intensities are monitored and analyzed in terms of its mechanistic implications.

1590-Pos Board B434**Fabrication of SERS Active Gold Nanoarrays for the Study of Adsorbed Phospholipids**

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In this study Surface-enhanced Raman scattering (SERS) was employed to study phospholipid molecules adsorbed on nanometre-sized Au structures. Highly ordered gold nano-rod arrays were produced using anodized aluminum oxide templates and electrochemical deposition. The nanorods were imaged with scanning electron microscopy and had an aspect ratio of 10. Raman spectra of 4-aminothiophenol on the prepared substrates showed an enhancement of 102 when compared with a smooth gold surface. Lipids were deposited on the nanoarrays and peaks at 2850 and 2927 cm⁻¹ in the Raman spectra were identified as CH₂ and CH₃ stretches respectively. It has been demonstrated that nano-rod textured electrodes are excellent substrate for SERS of biomolecules at metal surfaces.

1591-Pos Board B435**FTIR Spectroscopic Studies on Protein Migration in the Retina due to Light Exposure**Diana E. Bedolla Orozco¹, Lisa Vaccari², Vincent Torre¹.¹SISSA, Trieste, Italy, ²Sincrotrone Trieste S.C.p.A., Trieste, Italy.

Phototransduction, the initial event in vision, starts in rod and cone photoreceptors and continues across the entire retina. Some of the chemical processes that are triggered by light involve the migration of proteins to and from distinct compartments of the photoreceptors cells. In this work, we analyzed the protein migration process by imaging thin mouse retina slices using FTIR spectromicroscopy, both with conventional and Synchrotron Radiation (SR) source. We studied protein migration with IR spectroscopy, since it is a label-free technique, not dependent on antibodies of specific proteins. By using a Focal Plane Array (FPA) IR-bidimensional detector coupled to a FTIR microscope, we acquired in one shot 4096 spectra, from which we obtained the distribution of chemical components. From the analysis of Amide bands, we identified the protein location following light exposure of increasing duration and followed their translocation. We used SR from the light source of ELETTRA in Trieste (Italy) to obtain better quality spectra at the same spatial resolution from specific regions of the retina.

1592-Pos Board B436**Determination of Simvastatin Induced Variations in Sciatic Nerve by ATR-FTIR Spectroscopy**Kumsal Ozgun¹, Nihal Simsek Ozek², Feride Severcan².¹Bilkent University, Department of Molecular Biology and Genetic, Ankara, Turkey, ²Middle East Technical University, Department of Biology, Ankara, Turkey.

Simvastatin, a lipophilic statin derived drug, is commonly used drug for treating lipid and cardiovascular disorders. It is well known that the long term usage of this drug leads to peripheral neuropathy, mononeuropathy and memory prob-

lems. Moreover a possible treating agent in central nervous system disease like Alzheimer has been reported. The results that were obtained from simvastatin and nervous system are clinically unclear and controversial. Therefore the current study was aimed to clarify the possible effects of simvastatin on the content, composition, dynamics and conformation of macromolecules in rat sciatic nerve using ATR-FTIR Spectroscopy. Experimental male adult rats were divided two groups as control (n=10) and simvastatin-treated group (n=10). 50 mg simvastatin/ kg was given to treated group by oral gavage for 1 month. In the FTIR spectra, the shift in peak positions, the change in bandwidths and the intensity/area values of the bands were determined and compared in between control and treated groups. The results revealed that there is a significant decrease in amount of unsaturated lipid in simvastatin treated sciatic nerve, which indicates an increase in lipid peroxidation. Moreover, with simvastatin treatment a reduction in the saturated lipid, protein, glycogen and nucleic acid content was found showing degradation or decrease in the synthesis of these molecules. Protein secondary structure was found to be changed as a decrease in α -helix and β -sheet content and an increase aggregated β -sheet and random coil content in treated samples implying protein denaturation. The outcomes of this study show that high dose simvastatin application leads to structural and molecular variations in sciatic nerve by affecting its macromolecular composition.

Keywords: sciatic nerve, simvastatin, statin, ATR-FTIR spectroscopy, protein secondary structure.

1593-Pos Board B437**Using Difference Infrared Spectroscopy to Investigate the Effects of pH on PGK-Substrate Complexes**

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Yeast phosphoglycerate kinase catalyzes the reversible phosphate transfer in the reaction: ADP + 1,3-bis-phosphoglycerate \leftrightarrow ATP + 3-phosphoglycerate. Prior research indicates a hinge-bending mechanism occurs during catalysis to bring the substrates into closer proximity. Domain closure is only initiated in ternary complexes, in which both substrates are simultaneously bound to the enzyme. The activity and conformation of PGK is directly influenced by substrate and salt concentrations as well as pH. For example, activity assays confirm that PGK activity increases from pH 6.5 to 7.5. To determine the effects of pH on the conformational changes of PGK, we used difference Fourier transform infrared spectroscopy (FTIR) in conjunction with caged nucleotides. Difference infrared data associated with nucleotide (ATP or ADP) binding to PGK or PGK-3PG complexes was compared at pH 5.5, 6.5 and pH 7.5. Circular dichroism was also used to study PGK secondary structure at the aforementioned pH conditions. Comparison of the difference FTIR data allowed the isolation of pH dependent vibrations that arise from protein conformational changes induced by substrate binding. We have identified multiple vibrations that are associated with the PGK ternary complex and are influenced by pH. Difference FTIR studies resulted in the identification of specific changes within amino acid side chains and protein secondary structures that are altered by pH and associated with ternary complex formation.

1594-Pos Board B438**Resolving the Complications Induced by the Side Chain Carbonyl of Glutamine**

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IR spectroscopy is a widely used technique for structural studies of proteins which focuses on the amide I mode, an efficient marker of secondary structure. However, the use of IR for studying the glutamine based proteins associated with diseases such as Huntington's disease and Spinobulbar muscular atrophy is limited due to the additional complications introduced by the side chain carbonyl of glutamine. The two carbonyls will both vibrate in the amide I region, giving rise to multiple amide I modes; these must be resolved and unequivocally assigned if one is to understand the IR spectra of proteins containing glutamine. Furthermore, there is the opportunity for coupling between the two carbonyls through an electrostatic interaction known as transition dipole coupling (TDC). TDC causes a delocalization of the amide I vibrational wavefunction across oscillators that vibrate at similar frequencies (e.g. the backbone carbonyls of a protein or the two carbonyls of glutamine) and affects the frequency and transition strengths of the observed IR bands. TDC is dependent on the proximity and alignment of the carbonyls and thus is affected by the secondary structure of the protein as well as the side chain arrangement.

Here we report a fundamental study of the IR spectra of glutamine that will ultimately guide the spectral assignment of the many amide I bands observed in the IR spectra of long stretches of polyglutamine residues (pQ32 and pQ43). We explore the hypothesis that the side chain of glutamine can form a hydrogen

bond with the backbone, resulting in a cyclic conformation with a fixed alignment of the two carbonyls. The amidated form of glutamine has been studied at varying pH and temperature; data has been compared to that obtained for amidated alanine, amidated asparagine and non-amidated glutamine.

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Chemical Exchange 2DIR of Base Pair Opening Fluctuations in RNA Tetraloops: A Simulation Study

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Hydrogen bonds play an important role in RNA structure and dynamics. Fluctuations in base pair openings could be the starting point of unfolding processes or could indicate potential docking sites for ligands or proteins. The effects of hydrogen bond formation and breaking kinetics in RNA base pairs on the linear and coherent third order infrared spectra of small UUCG tetraloops in solution can be described by Markovian, not necessarily Gaussian, fluctuations. We have simulated these spectra using the stochastic Liouville equations. Slow fluctuations are described phenomenologically. Fast fluctuations are characterized by an N-state jump model for hydrogen bond configurations, where N depends on the specific tetraloop. Bases in the RNA strands that exhibit high levels of fluctuation are isotope labeled and the chemical exchange 2DIR spectra are calculated. The existence and evolution of the resultant cross peaks at different waiting times provides information on the coupling interactions between base pairs in the loops, which will be used to help characterize unfolding mechanisms of the RNA strands.

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Nitrile-Modified Nucleosides as a Probe of Local Nucleic Acid Environments

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The nitrile functional group has been extensively investigated as a probe in proteins but not as a probe in nucleosides. The solvent-induced vibrational frequency shift of the nitrile band of 5-cyano-2'-deoxyuridine was examined in THF-H₂O mixtures. The nitrile stretching frequency ($\nu_{C\equiv N}$) exhibited moderate solvent sensitivity, undergoing a 9.2 cm⁻¹ blue shift from THF to H₂O, and varied linearly with temperature, exhibiting a 1.4 cm⁻¹ red shift from 290 K to 340 K in H₂O. The $\nu_{C\equiv N}$ of 5-cyano-2'-deoxy-3',5'-bis-O-(*t*-BuPh₂Si)-uridine underwent a 1.3 cm⁻¹ blue shift when titrated with a base-pairing mimic, 2,6-diheptanamido-pyridine to yield an association constant of 90 M⁻¹. The corresponding C¹⁵N labeled nucleoside is currently under investigation by ¹⁵N NMR to determine the utility of the C¹⁵N moiety as an NMR probe of the local environment of nucleic acids. Both the IR and NMR results will be presented complemented by density functional theory calculations.

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Multidimensional Optical Spectroscopy Of Proteins Out Of Thermal Equilibrium

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In recent years, nonlinear multidimensional optical spectroscopy has been used as a highly sensitive probe of molecular dynamics in the condensed phase. Multidimensional optical spectroscopy builds upon the methodology of two-dimensional nuclear magnetic resonance spectroscopy and applies the same principles to vibrational and electronic resonances such that these techniques may be used as an ultrafast probe of molecular dynamics. In particular, these techniques have been used to study the thermal unfolding of proteins following a nanosecond temperature jump. In this study, we examine the multidimensional optical spectra of several biological systems of interest out of thermal equilibrium by using molecular dynamics to develop snapshots of the systems and the SPECTRON software package to calculate the spectroscopic signals. In order to enhance conformational sampling, an artificial temperature is used; the exact correlation functions of the system contributing to the material response are recovered using an action-reweighting scheme based on a stochastic path-integral formalism. The calculated spectra provide information on the states sampled by the system during the course of thermal unfolding.

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Spatially-resolved Analysis Of DNA Nanocomplex Self-assembly Enabled By Integrating Nanophotonics And Microfluidics

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Advances in genomics continue to fuel the development of future therapeutics that can target pathogenesis at the cellular and molecular level. Often functional only inside the cell, nucleic acid-based therapeutics require an efficient intracellular delivery system. One widely adopted approach is to complex DNA with a gene carrier to form nanocomplexes via electrostatic self-assembly, facilitating cellular uptake of DNA while protecting it against degradation. The challenge, however, lies in rational design of gene carriers, since premature dissociation or overly stable binding would be detrimental to the cellular uptake and therapeutic efficacy. Nanocomplexes synthesized by bulk mixing showed a diverse range of intracellular unpacking and trafficking behavior, which was attributed to the heterogeneity in size and stability of nanocomplexes. The heterogeneity of nanocomplexes resulting from bulk synthesis hinders the accurate assessment of the self-assembly kinetics and adds to the difficulty in correlating their physical properties to transfection efficiencies or bioactivities. We present a novel convergence of nanophotonics (i.e. QD-FRET) and microfluidics to characterize kinetic aspect of the nanocomplexes synthesis under laminar flow in real-time. QD-FRET provides a highly sensitive and quantitative indication of the onset of molecular interactions and throughout the process of nanocomplexes synthesis, whereas microfluidics offers a well-controlled microenvironment to spatially analyze the process with high temporal resolution (~milliseconds). For the model system of polymeric nanocomplexes, two distinct stages in the self-assembly process were captured by this analytic platform. The kinetic aspect of the self-assembly process obtained at the microscale would be particularly valuable for microreactor-based reactions which are relevant to many micro- and nano-scale applications. Further, customized nanocomplexes may be generated through proper design of microfluidic devices, and the resulting QD-FRET polymeric DNA nanocomplexes could be readily applied for establishing structure-function relationships.

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Oligonucleotide Microarray Analysis with Single Molecule Sensitivity

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We present a microarray analysis platform, which enables detection of hybridized DNA sequences at the level of single molecules. The readout is performed on a high sensitivity chip scanner based on an fluorescence microscope. Capture sequences were printed on custom-made aldehyde-functionalized glass coverslips. The microarray performance was tested with a 60mer fluorescent oligonucleotide hybridized to its complementary sequence, immobilized on the biochip. The determined dynamic range of the platform reaches 4.7 orders of magnitude with a sensitivity of 1.3 fM. Furthermore mRNA expression profiling experiments of tetracycline (un)treated HaCat cells were performed. For such competitive hybridization experiments only 5% reverse transcribed cDNA out of 5 µg total RNA were hybridized, a hundredfold lower amount than used typically for commercial microarrays. Such wide range in detection sensitivity needs reliable methods for exact data quantification. At low concentration the signal of each spot and molecule brightness was quantified by counting the molecules, fitting them with a 2-dimensional Gaussian function. For high concentrations, the number of molecules per spot was inferred from the total signal per spot. Good correlation of the data with experiments on commercial microarrays using hundredfold higher sample amounts indicates the feasibility of this approach, which avoids application of error prone amplification methods.

References

- 1.) Jaroslav Jacak, Jan Hesse, Maria Kasper, Fritz Aberger, Annemarie Frischauf, Stefan Howorka, and Gerhard J. Schütz - Proc.SPIE, 5699(2005), 442-449.
- 2.) Hesse, J., Sonnleitner, M., Sonnleitner, A., Freudenthaler, G., Jacak, J., Hoglinger, O., Schindler, H., Schutz, G.J. - Analytical Chemistry, 76 (2004), 5960-5964.
- 3.) J. Hesse, J. Jacak, M. Kasper, G. Regl, T. Eichberger, M. Winklmayr, F. Aberger, M. Sonnleitner, R. Schlapak, S. Howorka, L. Muresan, Anna-Maria Frischauf, Gerhard J. Schütz - Genome Res., 2006, 16, 1041-45.

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Viscosity Measurement of Biological Fluids Using Optical Tweezer

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